

# Measures of disease frequency and association

## Standard errors and confidence intervals

### 1 Illustrative dataset

To illustrate the estimation of the measures of disease frequency, associated standard errors (se) and confidence intervals (CI), we will use the BrCa dataset from the Epi package:

```
library(Epi)
data(BrCa, package = "Epi")
## only consider some of the variables
BrCaR <- BrCa[,c("pid","age","grade","tox","xst")]
## give more intuitive name
names(BrCaR) <- c("id","age","grade","time","status")
## display
str(BrCaR)
```

```
'data.frame':      2982 obs. of  5 variables:
 $ id      : int  1264 1150 838 1214 1130 1118 386 1417 927 489 ...
 $ age     : int   54 55 34 42 35 50 46 40 36 42 ...
 $ grade   : Factor w/ 2 levels "2","3": 1 1 1 1 1 1 1 1 1 1 ...
 $ time    : num  12.97 8.78 9.41 10.47 10.35 ...
 $ status  : Factor w/ 2 levels "Alive","Dead": 1 1 1 1 1 2 1 1 1 1 ...
```

It contains  $n = 2982$  women with breast cancer (with grade indicated by the variable `grade`) followed from diagnosis until death or loss to follow-up. Summing `time` and `status` values over participants gives the number of events and total follow-up time:

```
c("riskTime" = sum(BrCaR$time), "person.year" = sum(BrCaR$status=="Dead"))
```

```
riskTime    event
21270.74    1272.00
```

The same calculation can be performed per group using `xtabs` (instead of subsetting the data set):

```
t23 <- xtabs(cbind(n=1,N. = status=="Dead",person.year = time) ~ grade, data = BrCaR)
t23
```

```
grade      n      N. person.year
 2    794.000    262.000    6323.439
 3    2188.000   1010.000   14947.300
```

## 2 Theory

Denote by:

- $n$  the number of persons in the population under study.
- $H_{\bullet}(t)$  the number of persons sick at time  $t$ .
- $N_{\bullet}(t)$  the number of persons who contracted the disease at some point between time 0 and time  $t$ .
- $\int_0^t Y_{\bullet}(s)ds = \sum_{i=1}^n \min(T_i, t)$  the cumulative at risk time up to time  $t$  (**person.year** in the example dataset). For a given subject, it is the time during which he is under study and has not yet contracted the disease.

statistic	estimate	standard error (se)	confidence interval [lower, upper]
prevalence	$\hat{\pi}(t) = \frac{H_{\bullet}(t)}{n}$	$\hat{\sigma}_{\hat{\pi}(t)} = \sqrt{\frac{\hat{\pi}(t)(1-\hat{\pi}(t))}{n}}$	lower= $\hat{\pi}(t) - 1.96\hat{\sigma}_{\hat{\pi}(t)}$ upper= $\hat{\pi}(t) + 1.96\hat{\sigma}_{\hat{\pi}(t)}$
odds	$\hat{\Omega}(t) = \frac{H_{\bullet}(t)}{n-H_{\bullet}(t)}$	$\sigma_{\log \hat{\Omega}(t)} = \sqrt{\frac{1}{H_{\bullet}(t)} + \frac{1}{n-H_{\bullet}(t)}}$	lower= $\hat{\Omega}(t) \exp(-1.96 \sigma_{\log \hat{\Omega}(t)})$ upper= $\hat{\Omega}(t) \exp(1.96 \sigma_{\log \hat{\Omega}(t)})$
incidence rate	$\hat{\lambda}(t) = \frac{N_{\bullet}(t)}{\int_0^t Y_{\bullet}(s)ds}$	$\sigma_{\log \hat{\lambda}(t)} = \frac{1}{\sqrt{N_{\bullet}(t)}}$	lower= $\hat{\lambda}(t) \exp(-1.96\sigma_{\log \hat{\lambda}(t)})$ upper= $\hat{\lambda}(t) \exp(1.96\sigma_{\log \hat{\lambda}(t)})$
risk	$\hat{r}(t) = \frac{N_{\bullet}(t)}{n}$	$\hat{\sigma}_{\hat{r}(t)} = \sqrt{\frac{\hat{r}(t)(1-\hat{r}(t))}{n}}$	lower= $\hat{r}(t) - 1.96\hat{\sigma}_{\hat{r}(t)}$ upper= $\hat{r}(t) + 1.96\hat{\sigma}_{\hat{r}(t)}$

When contrasting two groups composed of distinct individuals, the standard error of the difference is the square root of the sum of squared standard errors. For instance if we estimate:

- a risk  $\hat{r}_1(t)$  based on  $n_1$  persons in one group, with standard error  $\hat{\sigma}_{\hat{r}_1(t)} = \sqrt{\frac{\hat{r}_1(t)(1-\hat{r}_1(t))}{n_1}}$
- a risk  $\hat{r}_2(t)$  based on  $n_2$  persons in the other group, with standard error  $\hat{\sigma}_{\hat{r}_2(t)} = \sqrt{\frac{\hat{r}_2(t)(1-\hat{r}_2(t))}{n_2}}$

The standard error for the risk difference can be estimate with  $\hat{\sigma}_{\hat{r}_2(t)-\hat{r}_1(t)} = \sqrt{\frac{\hat{r}_2(t)(1-\hat{r}_2(t))}{n_2} + \frac{\hat{r}_1(t)(1-\hat{r}_1(t))}{n_1}}$ .

## 3 Implementation

### 3.1 'by hand': using the entire follow-up

Consider the table `t23` previously created which contains the number of women (`n`), events (`N.`), and at risk time (`person.year`) for each group (grade 2 and grade 3). We can add an additional line for the overall cohort by adding the group specific values using `addmargin`:

```
t33 <- addmargins(t23, margin = 1) ## 1: sum over rows, 2 over columns
t33
```

grade	n	N.	person.year
2	794.000	262.000	6323.439
3	2188.000	1010.000	14947.300
Sum	2982.000	1272.000	21270.738

Then we can estimate the incidence rates (in each group and for the whole cohort) as:

```
lambda <- t33[,"N."]/t33[,"person.year"]
unnname(lambda)
```

```
[1] 0.04143315 0.06757073 0.05980046
```

Confidence intervals can then be obtained using:

```
se.loglambda <- 1/sqrt(t33[,"N."])
cbind(estimate = lambda,
      lower = lambda * exp(-1.96 * se.loglambda), upper = lambda * exp(1.96 * se.loglambda))
```

	estimate	lower	upper
2	0.04143315	0.03670790	0.04676666
3	0.06757073	0.06352934	0.07186921
Sum	0.05980046	0.05660276	0.06317882

Having incomplete follow-up for many women

```
quantile(BrCaR$time[BrCaR$status=="Alive"])
```

	0%	25%	50%	75%	100%
	0.09856263	7.01437394	8.81314150	10.60780271	19.28268305

complicates the evaluation of the risk. We could use the risk-rate relationship to evaluate the 2 year risk (in each group and for the whole cohort):

```
1 - exp(-lambda * 2)
```

	2	3	Sum
	0.0795258	0.1264077	0.1127255

Both the estimation of the rate and of the risk assume that the rate is constant over the entire follow period (about 20 years).

## 3.2 'by hand': on a restricted follow-up time

Consider only the first 2 years of follow-up. We can re-compute the summary statistics (number of individual, number of events, total follow-up time) either by adding new columns to the dataset:

```
BrCaR$time2 <- pmin(BrCaR$time,2)
BrCaR$status2 <- ifelse(BrCaR$time<=2,as.character(BrCaR$status),"Alive")
xtabs(cbind(n=1,N. = status2=="Dead", person.year = time2) ~ grade, data = BrCaR)
```

or directly do the modification in xtabs:

```
t23.y2 <- xtabs(cbind(n=1,
                    N. = (status=="Dead")*(time<=2),
                    person.year = pmin(time,2)) ~ grade, data = BrCaR)
t33.y2 <- addmargins(t23.y2, margin = 1)
t33.y2
```

grade	n	N.	person.year
2	794.000	27.000	1563.507
3	2188.000	191.000	4229.844
Sum	2982.000	218.000	5793.351

We can evaluate the incidence rate and 2 year risk as:

```
lambda.y2 <- t33.y2[,"N."]/t33.y2[,"person.year"]
rbind(rate = lambda.y2, risk = 1 - exp(- lambda.y2 * 2))
```

	2	3	Sum
rate	0.01726887	0.04515533	0.03762934
risk	0.03394812	0.08635269	0.07249648

Compared to the estimation based on the entire follow-up, we use a weaker assumption (constant rate within the first two years only) but have less events to work with (i.e. larger statistical uncertainty):

```
se.loglambda.y2 <- 1/sqrt(t33.y2[,"N."])
cbind(estimate = lambda.y2,
      lower = lambda.y2 * exp(-1.96 * se.loglambda.y2),
      upper = lambda.y2 * exp(1.96 * se.loglambda.y2))
```

	estimate	lower	upper
2	0.01726887	0.01184260	0.02518145
3	0.04515533	0.03918475	0.05203564
Sum	0.03762934	0.03295148	0.04297128

Note that had we have had no loss to follow-up we could have computed the 2 year risk without making assumptions on the incidence rate doing:

```
t33.y2[,"N."]/t33.y2[,"n"]
```

	2	3	Sum
	0.03400504	0.08729433	0.07310530

### 3.3 'glm': on a restricted follow-up time

The `glm` function can be used with the `poisson` family to estimate incidence rates:

```
e.pois <- glm(status2=="Dead" ~ grade, offset = log(time2),
             family = poisson(link="log"), data = BrCaR)
cbind(estimate = exp(coef(e.pois)), exp(confint(e.pois)))
```

Waiting for profiling to be done...

```
              estimate      2.5 %      97.5 %
(Intercept) 0.01726887 0.01154676 0.02462553
grade3      2.61484005 1.78076822 3.99981206
```

provides the incidence rate for the reference group (here `grade2`) and the rate ratio (`grade3` 3 vs. 2). Note that removing the intercept in the formula leads to the same model:

```
e.pois2 <- glm(status2=="Dead" ~ grade-1, offset = log(time2),
              family = poisson(link="log"), data = BrCaR)
logLik(e.pois2)
logLik(e.pois)
```

```
'log Lik.' -881.9157 (df=2)
```

```
'log Lik.' -881.9157 (df=2)
```

but parametrized differently: an incidence rate per group

```
cbind(estimate = exp(coef(e.pois2)), exp(confint(e.pois2)))
```

Waiting for profiling to be done...

```
              estimate      2.5 %      97.5 %
grade2 0.01726887 0.01154676 0.02462553
grade3 0.04515533 0.03905040 0.05186517
```

The incidence rate for the whole cohort can be obtained by:

```
e.pois <- glm(status2=="Dead" ~ 1, offset = log(time2),
             family = poisson(link="log"), data = BrCaR)
c(exp(coef(e.pois)), exp(confint(e.pois)))
```

Waiting for profiling to be done...

```
(Intercept)      2.5 %      97.5 %
0.03762934 0.03285262 0.04284772
```

The CIs are slightly different as they are based on a different approximation (profile likelihood instead of delta method).

For the risk ( $\Delta$  in absence of censoring) one can use the `binomial` family:

- with an identity link to get risk difference (no back-transformation)

```
e.RD <- glm(status2=="Dead" ~ grade, family = binomial(link="identity"), data = BrCaR)
cbind(coef(e.RD), confint(e.RD))
```

Waiting for profiling to be done...

```
                2.5 %    97.5 %
(Intercept) 0.03400504 0.02286881 0.04814043
grade3      0.05328929 0.03530580 0.07012383
```

- with an log link to get risk ratio (back-transformation via `exp`)

```
e.RR <- glm(status2=="Dead" ~ grade, family = binomial(link="log"), data = BrCaR)
cbind(exp(coef(e.RR)), exp(confint(e.RR)))
```

Waiting for profiling to be done...

```
                2.5 %    97.5 %
(Intercept) 0.03400504 0.02286695 0.04813859
grade3      2.56709984 1.76421904 3.89779150
```